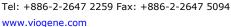
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User Bulletin

Plant Genomic DNA Extraction Miniprep System

Isolation of genomic DNA from 100 mg plant material or 1×10^8 cells.

Downstream Application

- * Restriction digestion
- * Southern Blotting
- * RAPD, RFLP
- * PCR, Real-Time PCR

Product Contents

GPG1001	GPG1002
50	250
24ml	120ml
8ml	40ml
18ml	90ml
20mg	110mg
15ml	45ml x 2
50	250
50	250
100	500
1	1
	24ml 8ml 18ml 20mg 15ml 50 50

All buffers need to be mixed well before use.

Shipping & Storage

Viogene Plant Genomic DNA Extraction Miniprep is shipping and storage at ambient temperature up to 12 months.

If precipitate form by freezing temperature on any buffer, warm up at 37°C to redissolve.

Protocol

❖ Please read the following notes before starting the procedures.

Important Notes

- All centrifugation should be done at room temperature.
- Preheat a water bath to 65°C.
- Preheat TE or ddH₂O to 65°C for DNA elution.
- PX1 buffer may form unclear upon storage. This does not affect the procedure and efficiency.
- PX1 Buffer may form a precipitate, warm at 65°C to redissolve.

For GPG1001

- Add 200 μ I of ddH₂O to the RNase A powder tube, vortex to dissolve and store at 4° C.
- Add 60 ml of ethanol (98~100 %) to the WS Buffer bottle when first open the bottle.

For GPG1002

- Add 1100 µ I of ddH₂O to the RNase A powder tube, vortex to dissolve and store at 4° C.
- Add 180 ml of ethanol (98~100 %) to the WS Buffer bottle when first open the bottle.

Viogene's unique design — EasyLid™

The $\mathsf{EasyLid}^\mathsf{TM}$ is designed to prevent contamination during the procedure.

Tips for EasyLidTM –

Twist the arm of the cap and pull the cap to break the EasyLidTM.



1. Grind 100 mg (or less) plant sample under liquid nitrogen to fine powder and transfer to a new tube.

Do not allow the sample to thaw, and continue immediately to step 2.

2. Add 400 μ l of PX1 Buffer and 4 μ l of RNase A solution (100 mg/ml) to the tissue powder and vortex vigorously, then incubate the mixture at 65 $^{\circ}$ C for 10 minutes.

- 3. Add 130 μ l of PX2 Buffer to the lysate, vortex, and incubate on ice for 5 minutes.
- 4. Apply lysate to the Shearing Tube sitting in a Collection Tube and centrifuge at full speed for 2 minutes. Transfer flow-through sample from the Collection Tube to a new tube.

Avoid pipetting any debris or pellet in the collection tube.

5. Add 0.5 volume of PX3 Buffer and 1 volume of 98-100% ethanol to the clear lysate and invert the tube $3 \sim 5$ times.

For example: If 450 μl clear lysate collected, add 225 μl PX3 Buffer and 450 μl ethanol.

6. Apply 650 μ l of the ethanol added sample (including any precipitate) from step 5 to a Plant Genomic DNA Mini Column sitting in a Collection Tube, close the cap, centrifuge at 8,000 x g (10,000 rpm) for 1 minute, and discard the filtrate.

If the solution remains above the membrane, centrifuge again at 13,000 rpm.

- 7. Repeat step 6 for rest of the sample.
- 8 Wash the column twice with 0.7 ml of WS Buffer by centrifuging at full speed (13,000 rpm or 10,000 x g) for 30 \sim 60 seconds and discard the filtrate.

Add ethanol (see Important Notes) to the WS Buffer bottle when first open the bottle.

- Centrifuge at full speed for 3 minutes to remove traces of WS Buffer.
- 10. Transfer the column to a new 1.5 ml tube, add 200 μ l of 65°C TE or ddH₂O, Stand the column for 5 minutes, and centrifuge for 1-2 minutes to elute DNA.
- 11. Centrifuge at full speed for 1 minute to elute DNA.
- 12. Store DNA at -20℃.